

R b	b	bbP a a b dM ba b
A b 7 4 1	P b 7 O ab M b b b b	
A b ba7 July 23 rd , 2015	Ob 7 3	M db 7 1

Queen's University

Principal Investigators, Research Staff, Veterinary Staff

R	b		b	bbP	a	a	b	dM	ba	b
A	b	7		P	b	7				
		41		O	ab	M	b	b	b	b
A	b	ba7		Ob	7			Mdb	7	
		July 23 rd , 2015			3				2	

Nutra-gel or moist chow

R	b	b	bbP	a	a	b	dM	ba	b
A	b	7	P	b	7	O	ab	M	b
	4	1		b	b	b	b	b	b
A	b	ba7	Ob	7		M	db	7	
		July 23 rd , 2015		3				3	

or blue-tinged indicate distress).

- o Color and moisture of the mucous membranes – these should be pink and moist with a capillary refill time of < 2 sec.
- o Temperature: palpate the extremities (feet or tail) and/or take a rectal temperature to ensure that temperature does not go outside of physiologic parameters.

Until the blinking reflex has returned, ensure eye lubricant is instilled.

If recovery is >5 minutes turn animal every 5 minutes. This stimulates the animal and also alternates the side of the chest (and lung) which the animal is laying on, thereby facilitating respiration.

Put a measured amount of food or Nutra-gel at the bottom of the cage.

Alternatively, moist chow can be provided during the post-operative period to encourage appetite and facilitate hydration. Nutra-gel must be changed daily.

Moist chow: fill a small bowl or petri dish ¾ full with warm water and top up with a few (2-3) food pellets. Let sit for 10 minutes to allow water to be absorbed before placing in the animals cage.

Animals should only be returned to an animal room or to their cage mates once they have recovered their righting reflex and are ambulatory.

Examine animal daily for signs of grooming, general appearance, posture and locomotor activity. If any signs of dehydration occur (ruffled coat, sunken eyes, prolonged skin tenting), administer 30-60 ml/kg of sterile Lactated Ringer's solution or sterile 0.9% sodium chloride SC daily until the animal's condition improves.

Ensure the animal is eating, drinking and eliminating normally.

Measure and record post-operative body weight daily, for a minimum of three days.

All procedures and drugs must be recorded on cage cards. Surgical sheets or forms are recommended, easily accessible by the Veterinary team (e.g. stored in the anteroom or treatment room). However, if all the information found in the book is recorded on the cage card, the Principal Investigator can forego this secondary documentation. General parameters to be recorded for rodent surgery includes: Surgery (on front of cage card), baseline and post-op Day 1, 2, 3 weight, anaesthetics (dose, rate, frequency), all pre-operative and post-operative analgesic and fluids (dose, rate, frequency).

Any irregularities in recovery must be brought to the attention of the clinical veterinarian. Continued weight loss, dehydration and lethargy are NOT acceptable. Refer to the Humane Interventions within the Animal Use Protocol (and the veterinarians) for guidance.

Monitor the animal daily for signs of surgical complications such as (but not limited to) pain, paralysis, or seizures.

R	b		b	bb	P	a	a	b	d	M	ba	b
A	b	7			P	b	7					
		41			O	ab	M	b	b	b	b	
A	b	ba7			Ob	7			M	db	7	
		July 23 rd , 2015				3					5	

SX LOG: STUDENT AND LAB NAME: _____ CONTACT #: _____ PROTOCOL #: _____

3-4 Seed pellets SC flasks? (1) SUBJECT

DAY 1 POST

Day 2 POST

DAY 3 POST

Medium Analgensis (name)

Drinking Analgensis (name)

NOISE RECOVERY STAIRS - EAR2, URINE, REDS, Analgensis (name)

3-4 Seed pellets SC flasks? (1) SUBJECT

DAY 1 POST

Day 2 POST

DAY 3 POST

3-4 Seed pellets SC flasks? (1) SUBJECT

DAY 1 POST

Day 2 POST

DAY 3 POST