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Nosecone

Isoflurane

Ketamine 100mg/kg (controlled substance – exemption permit required)

Xylazine (20mg/mL)

Acepromazine (10mg/mL)

Medetomidine

Atipamezole

Sterile sodium chloride 0.9% or

Sterile water (diluent)

Lactated Ringers solution (fluid therapy)

Sterile needles (various sizes)

Alcohol swabs

Gauze

Eye Lubricant

10mL red top vacutainers

3. Procedures:

:

i. Induction

- 1. Place the animal in the induction chamber.
- 2. Adjust the oxygen flow to 1 1.5 L/min.
- 3. Adjust the vaporiser to 3% 5%.
- 4. Once the mouse is in lateral recumbency and cannot right itself, apply eye lubricant using a sterile cotton tip applicator, and transfer the animal to a nosecone for maintenance.

ii. Maintenance

- 1. Ensure the mouse's snout is firmly seated in the nosecone with a good seal.
- 2. Adjust the oxygen flow to 800-1000 ml/min.
- 3. Adjust the vaporiser from 1 2.5% (this is variable depending on the depth of anesthesia required for the particular procedure or for the strain of the mouse).
- 4. Maintain the mouse on a heating device the duration of the procedure.

iii. Recovery

- 1. Turn off the isoflurane and leave the animal on 100% oxygen (short term).
- 2. Administer fluid therapy.
- 3. Transfer the animal to a clean cage and place it on a piece of paper towel until it is mobile.
- 4. Provide heat during this recovery phase to half of the cage, to ensure that the mouse has the ability to escape the heat as necessary.
- 5. Return the animal to the colony room once it has fully recovered from anaesthesia (blinking and righting reflex have returned).

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(100ul) per 10g body weight IP.

- ${f v.}$ Duration of anaesthesia is approximately 30 minutes depending on the strain, sex, and body weight of the animal.
- vi. After 30 minutes, a half dose (0.05 ml per 10g) may be administered as needed.
- vii. Atipamezole (5 mg/ml) can be administered to reverse xylazine and facilitate recovery at a dose of 0.1 to 5.0 mg/kg SC or IP.

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Date	New Version
March 14, 2012	Created
December 15, 2016	Review and update
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